

Inhibition of the Type I Insulin-like Growth Factor Receptor Expression and Signaling: Novel Strategies for Antimetastatic Therapy

Pnina Brodt,* Amir Samani and Roya Navab

DEPARTMENTS OF SURGERY AND MEDICINE, MCGILL UNIVERSITY HEALTH CENTER, MONTREAL, QUEBEC, CANADA

ABSTRACT. The receptor for the type 1 insulin-like growth factor (IGF-1R) plays a critical role in the acquisition of the malignant phenotype. Using a highly metastatic murine lung carcinoma model, it was demonstrated that this receptor regulates several cellular functions that can impact on the metastatic potential of the cells, including cellular proliferation, anchorage-independent growth, cell migration, and invasion. The tumor model was used to develop several strategies for altering receptor expression and function as means of abrogating the metastatic potential of the cells. They include stable expression in the tumor cells of IGF-1R antisense RNA and dominant negative receptor mutants in which tyrosines in the kinase domain were substituted with phenylalanine. In addition, a novel strategy was used based on altering post ligand-binding receptor turnover. This led to inhibition of receptor re-expression and signaling and resulted in increased tumor cell apoptosis. When combined with the development of viral vectors designed to deliver genetic information with high efficiency, these strategies could form the basis for development of highly specific, antimetastatic therapy in tumors with known IGF-IR involvement. BIOCHEM PHARMACOL 60;8:1101–1107, 2000. © 2000 Elsevier Science Inc.

KEY WORDS. IGF-I receptor; cancer metastasis; signal transduction

The ability of cancer cells to metastasize and form cancerous lesions in secondary sites still poses the most formidable obstacle to cancer cure. Metastatic disease unresponsive to chemotherapy, surgery, and radiotherapy is fatal because it interferes with the functions of affected viscera such as liver, lung, bone, and brain. Lung and liver metastases, which are frequently associated with late-stage disease in malignancies such as breast and prostate carcinomas, carcinomas of the gastrointestinal tract, osteosarcoma, and melanoma, are generally refractory to common chemotherapeutic drugs and often inaccessible to surgical resection [1]. Therapeutic treatment of metastases in these vital organs will therefore require the development of innovative, biology-based approaches that can harness novel information on the molecular biology of metastasis to identify molecular targets and develop pharmaceutical reagents that can interfere with their expression and/or functions.

The establishment of a metastasis is the final outcome of a dynamic process that involves multiple interactions between the disseminating cancer cells and their rapidly changing microenvironments. It involves cell detachment from the primary tumor, migration, and invasion of host tissue barriers and intravasation. Once in the circulation, tumor cells may adhere to the vascular endothelium through specific receptors, extravasate into the target organ interstitium and parenchyma, and utilize both autocrine and paracrine growth regulatory mechanisms to grow in the secondary site. The newly formed lesions can themselves become the source of disseminating cells that repeat this cycle and give rise to tertiary metastases [2-4, see schematic representation in Fig. 1]. These multiple processes are mediated by a series of molecular interactions resulting from the deregulated expression and/or function of cell-cell and cell-extracellular matrix (ECM) adhesion receptors, ECM degrading proteinases, growth promoting factors, and their receptors [3]. Disruption of these molecular interactions at any one of these steps could potentially lead to abrogation of the entire process.

THE RECEPTOR FOR TYPE 1 INSULIN-LIKE GROWTH FACTOR (IGF-IR)

One family of molecules critical for malignant transformation and metastasis are the peptide growth factors that regulate cell entry into and progression through the cell cycle by binding to membrane receptor tyrosine kinases (RTK), which transmit signals to the nucleus through an intricate network of adaptor and signaling molecules [5–7]. One of the RTKs implicated in the induction and maintenance of the transformed/malignant phenotype is the re-

^{*} Corresponding author: Dr. Pnina Brodt, Dept. of Surgery, Division of Surgical Research, McGill University Health Center, Royal Victoria Hospital, Room H6.25, 687 Pine Ave W., Montreal, Quebec, Canada H3A 1A1. Tel. +1-514-842-1231; FAX +1-514-843-1411; E-mail: pnina.brodt@muhc.mcgill.ca.

1102 P. Brodt et al.

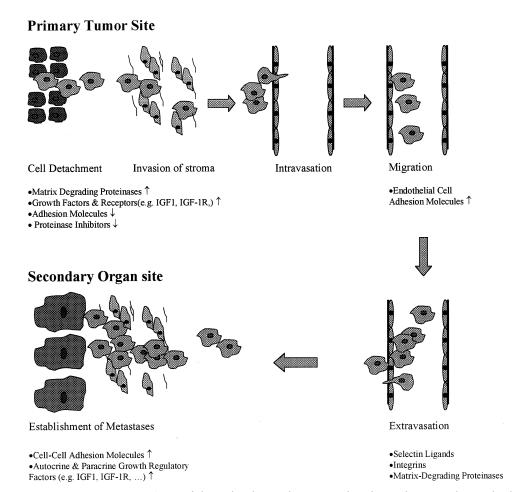


FIG. 1. The basic steps in cancer metastasis. Some of the molecular mechanisms and mediators known to be involved or altered at each of the steps in the metastatic cascade are indicated.

ceptor for the IGF-IR.* This heterotetrameric receptor consists of two 130-135 kDa α- and two 90-95 kDa β-chains, with several α-α and α-β disulfide bridges [8, 9]. The ligand-binding domain is located on the extracellular α -subunit. Approximately one-third of the β -subunit is extracellular and is connected to the intracellular portion by a single transmembrane domain. The intracellular region of the β-subunit has a binding site for phosphorylation substrates at tyrosine residue 950, an ATP-binding site at lysine 1003, a tyrosine kinase domain with 3 critical tyrosines at positions 1131, 1135, and 1136, and several tyrosines in the carboxyl domain at positions 1250, 1251, and 1316, all of which have been implicated in the regulation of the receptor's biological functions (Fig. 2). The IGF-IR ligands include IGF-I, IGF-II, and insulin, but it binds IGF-I, a 70-amino acid peptide with the highest affinity [10]. Binding and physiological activities of IGF-I can be modulated by its association with the IGF-binding proteins (IGFBPs), a family of structurally related, secreted proteins that bind both IGF-I and IGF-II with high affinities and regulate their biological accessibility and activity [11, 12].

The IGF-I receptor plays an important role in the regulation of the cell cycle [13]. During postnatal development and longitudinal growth, growth hormone functions are mediated via IGF-I. IGF-I serum levels are high during childhood, declining progressively after puberty. IGF-IR mRNA levels also decline considerably after puberty, remaining high in selected tissues such as the brain and kidney [10]. Malignant transformation, however, is often associated with up-regulated expression and/or constitutive activation of the IGF-IR [14]. A requirement for IGF-I for cell survival and growth has been documented in a broad range of cell types where it is thought to act in concert with initiation factors such as epidermal growth factor (EGF) and platelet-derived growth factor (PDGF) and mediate cell cycle progression from G_1 to S phase [13, 15, 16]. In addition to and distinct from its role as a positive regulator of cell growth, IGF-I is also a survival factor and could block apoptosis in a range of cell types in vitro [17, 18] and in vivo [19].

SIGNAL TRANSDUCTION BY THE IGF-IR

In response to ligand binding, the intrinsic tyrosine kinase of the receptor is activated, resulting in autophosphoryla-

^{*} Abbreviations: IGF-1R, receptor for the type I insulin-like growth factor; IRS, insulin receptor substrate; and PI-3K, phosphatidylinositide-3 kinase.

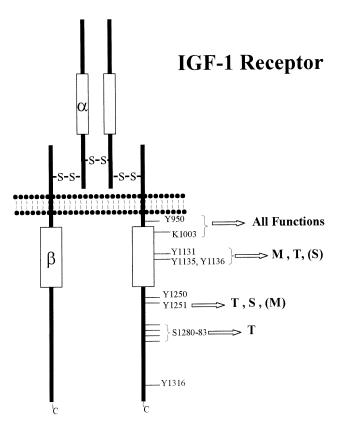


FIG. 2. Structure of the IGF-I receptor. Amino acid residues involved in signal transduction are shown. Their postulated roles in receptor-mediated functions based on mutational analyses are indicated on the right. M-mitogenic response, S-survival, T-transformation.

tion of tyrosines on the intracellular portion of the β -subunit and the subsequent tyrosine phosphorylation of several downstream substrates including the IRS 1-4 and Shc [20, 21]. The IRS constitute a family of structurally related adaptor proteins that can link the IGF-I receptor to downstream signal transduction mediators regulating cellular growth. Of these, IRS-1 is the most extensively studied [22]. This 165-195-kDa molecule does not contain SH2 (Src homology 2) or SH3 domains and may bind to the β-subunit through a PTB (pTyr-binding) domain [22, 23]. It contains at least 20 potential tyrosine phosphorylation sites and can act as a multisite "docking" protein associating with multiple downstream signaling proteins including PI-3 kinase [24], SH2 domain-containing tyrosine phosphatase (Syp) [25], Fyn, Nck, and growth factor receptor-bound protein-2 (Grb2) through their SH2 domains [26]. Stimulation of PI-3K leads to activation of several downstream substrates including protein kinase B (Akt), which can phosphorylate BAD and attenuate its proapoptotoic effect and the pp70 S6 kinase [20, 21]. Grb2 is tightly associated with the guanine nucleotide exchange factor mSOS linking the IGF-1R to the Ras/Raf-1/ mitogen-activated protein kinase (MAPK) signaling pathway, leading to activation of nuclear transcription factors [14, 27, see Fig. 3]. Like IRS-1, tyrosine phosphorylation of Shc promotes association with Grb2, linking it to the Ras pathway via the Grb2-mSOS complex [7]. Preferential phosphorylation of Shc or IRS-1 could depend on the cellular context and may direct IGF-IR signaling preferentially towards cellular proliferation or differentiation [28]. In some cell types, the IGF-IR can also directly phosphorylate the Janus kinases (JAK) 1 and 2 involved in cytokine-mediated signaling and JAK proteins may in turn phosphorylate IRS-1 [29]. Phosphorylation of JAK proteins can lead to phosphorylation/activation of signal transducers and activators of transcription (STAT) proteins, in particular STAT-3 [30], and STAT-3 activation may in turn be essential for the transforming activity of IGF-1R [31].

Other direct substrates of IGF-IR include the protooncogenes c-Crk II and CrkL and the p125 focal adhesion kinase (FAK) [32–34]. These molecules can link IGF-IR to integrin-mediated signaling and the cytoskeleton through p130(Cas) and paxillin, and the regulation of cell shape and motility [34, 35]. Because intracellular calcium levels increase in response to IGF-1 binding, PLC γ is also thought to be involved through its products inositol 1,4,5-triphosphate (IP₃) and 1,2-diacylglycerol (DAG) [14]. The relative importance of these pathways in signal transduction by IGF-IR is probably cell context-dependent and remains to be fully elucidated.

Mutational analyses have identified several domains in the receptor β-subunit containing amino acid residues essential for receptor functions. Lysine at position 1003 (the ATP-binding site) and tyrosine 950 (thought to be essential for IRS-1 and Shc binding and phosphorylation) are critical for all receptor functions [23, 36, 37]. Tyrosines 1131, 1135, and 1136 in the kinase domain are essential for the mitogenic and transforming activities of the receptor [38, 39], but there is some controversy regarding their role in regulating the antiapoptotic effect of the receptor [40]. In contrast, the tyrosines in position 1251, although not essential for receptor, IRS-1, and Shc phosphorylation, are critical for the transforming (anchorage-independent growth) and antiapoptotic effects of the receptor, possibly through involvement in regulating cytoskeletal reorganization [40-42]. The role of these tyrosines in mitogenesis is controversial [41, 42]. Finally carboxyl-terminal serines in positions 1280–1283 also appear to play a role in regulating the transforming function of the receptor [43, see Fig. 2].

ROLE OF IGF-1R IN MALIGNANCY

Several lines of evidence implicate IGF-1 and the receptor in malignant progression. Increased expression of IGF-I, IGF-IR, or both has been documented in many human malignancies including carcinomas of the lung, breast, thyroid, gastrointestinal tract and prostate, glioblastomas, neuroblastomas, rhabdomyosarcomas, and leukemias [reviewed in 44]. Furthermore, prospective clinical studies identified high plasma IGF-I levels as a potential risk factor for carcinomas of the breast, prostate, and colon [45–47]. In addition, the IGFs are potent mitogens for a wide range of tumor cell types *in vitro*. Several oncogenes have now been

P. Brodt et al.

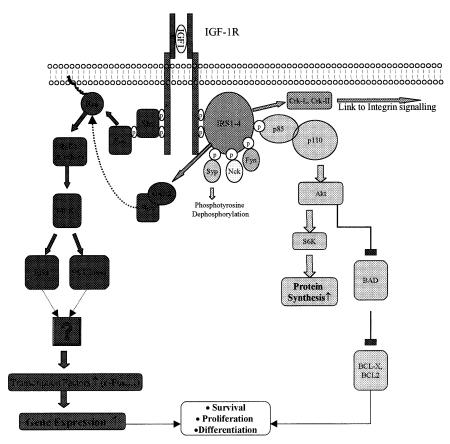


FIG. 3. Ligand binding-induced signal transduction by the IGF-I receptor. Ligand binding triggers IGF-I receptor autophosphorylation at multiple tyrosine residues. The intrinsic receptor tyrosine kinase then phosphorylates multiple substrates including IRS 1–4 and Shc. The IRS proteins act as docking sites for multiple proteins including PI-3K which can provide cell survival signals through activation of Akt. Shc and IRS phosphorylation links the receptor to the Grb-2/mSOS complex and the Ras/Raf-1/MAPK signaling cascade, leading to changes in gene transcription.

shown to affect IGF-I and IGF-IR expression [13]. Suppression of IGF-IR by antisense oligonucleotides [48], plasmids expressing IGF-IR antisense cDNA [49, 50], IGF-I peptide analogues [51], and triple-helix-forming oligodeoxynucleotides [52] suppressed tumor cell growth *in vivo* [reviewed in 53, 54].

ROLE OF IGF-IR IN METASTASIS

Our laboratory has been studying molecular factors that regulate tumor cell dissemination using a mouse tumor model of Lewis lung carcinoma sublines with distinct metastatic properties. IGF-IR was identified as a positive regulator of the invasive/metastatic phenotype and IGF-I as a paracrine growth-promoting factor in the liver [55, 56]. Highly metastatic carcinoma H-59 cells express high IGF-IR levels, and these were found to be critical for tumor cell ability to form metastases in the liver. H-59 cells expressing IGF-1R antisense RNA, lost mitogenic and motogenic responses to IGF-I, had reduced levels of the extracellular matrix-degrading metalloproteinase MMP-2, became non-invasive, and failed to form metastases in the liver following intrasplenic/portal inoculation [57, 58]. These studies identified IGF-IR as a potential biological

target for antimetastatic therapy and are the basis of ongoing efforts to develop additional antimetastatic strategies based on suppression of IGF-IR expression and function.

STRATEGIES FOR SUPPRESSION OF IGF-IR EXPRESSION AND FUNCTION

We investigated two additional approaches for suppression of IGF-IR expression and function: 1. Inhibition of signaling through the use of receptor dominant negative mutants and 2. Blocking receptor trafficking by inhibition of endosomal endopeptidases.

Use of Receptor Dominant Negative Mutants

Dominant negative mutants of the IGF-I receptor have been successfully used by other laboratories to block IGF-IR functions [59–60]. To investigate the potential use of dominant negative receptors to block tumor invasion and metastasis, we recently used site-directed mutagenesis to generate domain-specific mutants of the IGF-1 receptor β -subunit. Highly invasive IGF-IR $^+$ H-59 cells were transfected with a plasmid vector expressing full-length human

IGF-1 receptor cDNA in which the codons for tyrosines Y1131, Y1135, and Y1136 in the kinase domain were substituted with codons for phenylalanine. Stably transfected cells were analyzed with respect to changes in invasive and metastatic ability relative to unmodified cells or cells transfected with the wild-type receptor. The kinase domain mutant receptor had a dominant negative effect and significantly reduced the invasive and metastatic potential of these cells.* The results demonstrated that IGF-IR kinase domain mutants can block the metastatic potential of tumor cells in vivo, suggesting that dominant negative mutant receptors could provide an alternative or complementary approach to antisense-based reagents designed to reduce the number of functional receptors available on the cell surface. In a clinical setting, where micrometastases have either been diagnosed or are suspected, the successful use of such reagents will depend on the availability of highly efficient vehicles for delivery of genetic material into the tumor cells. Because normal liver parenchymal cells do not constitutively express measurable levels of IGF-IR and receptor levels are up-regulated in several malignancies with known hepatic involvement (e.g. colorectal and breast carcinomas), liver metastases may be particularly suited for IGF-IR-directed therapy based on the delivery of genetic information such as antisense or dominant negative receptors designed to alter receptor levels and/or functions.

Inhibitors of Receptor Trafficking

An alternative strategy for suppression of IGF-IR signaling is the targeting of post ligand binding events that regulate receptor turnover [36]. Like other peptide growth factors, binding of the IGF-1 to its receptor triggers receptormediated internalization of the ligand. This process is thought to be a key event in the regulation of receptor bioavailability and activity [62, 63]. IGF-I dissociates from its receptor within the acidic environment of the endosomes, a process that may be mediated by intraendosomal cysteine proteinases such as cathepsins B and L and is thought to be required for receptor recycling to the cell surface [62, 64-66]. Abrogation of ligand dissociation is known to lead to the intraendosomal accumulation of ligand and receptor and to disruption of receptor functions [63, 66]. Because this strategy only affects cycling cells, it is expected to have minimal deleterious effects on the majority of the normal, non-dividing hepatic cells, and therefore target cancer cells with high specificity. Recent reports including our own suggest that cysteine proteinase inhibitors can indeed alter tumor cell growth and IGF-IR signaling [67, 68].

CONCLUSIONS

The IGF-I receptor has been identified as a critical mediator of malignant transformation and a positive regulator of the metastatic phenotype. In recent years, it has emerged as a potential biological target for antimetastatic therapy. Several strategies for abrogation of IGF-IR expression and signaling have been tested in our model. They include manipulation of gene expression by antisense RNA, the use of dominant negative receptor mutants to alter receptor function, and inhibition of receptor—ligand processing to block receptor re-expression and signaling. Recent advances in the development of viral vectors for highly efficient delivery of genetic information into cells should provide the tools for translation of these experimental approaches into clinically relevant therapeutic modalities for the treatment of metastatic disease.

The authors are indebted to the Medical Research Council of Canada for support of this research.

References

- 1. Boring CC, Squires TS, Tong T and Montgomery S, Cancer Statistics, 1994. CA Cancer J Clin 448:7–26, 1994.
- Fidler IJ and Balch CM, The biology of cancer metastasis and implications for therapy. Curr Probl Surg 24:129–209, 1987.
- 3. Brodt P, Adhesion receptors and proteolytic mechanisms in cancer invasion and metastasis. *In: Cell Adhesion and Invasion in Cancer Metastasis* (Ed. Brodt P), pp. 167–242. R.G. Landes and Springer, Heidelberg and Austin TX, 1996.
- Liotta LA, Steeg PS and Stetler-Stevenson WG, Cancer metastasis and angiogenesis: An imbalance of positive and negative regulation. Cell 64:327–336, 1991.
- Pardee AB, G1 events and regulation of cell proliferation. Science 246:603–608, 1989.
- Yarden Y and Ullrich A, Growth factor receptor tyrosine kinases. Annu Rev Biochem 57:443–478, 1988.
- 7. Pawson T, Protein modules and signalling networks. *Nature* 373:573–580, 1995.
- 8. Ullrich A, Gray A, Tam AW, Yang-Feng T, Tsubokawa M, Collins C, Henzel W, Le Bon T, Kathuria S, Chen E, et al. Insulin-like growth factor I receptor primary structure: Comparison with insulin receptor suggests structural determinants that define functional specificity. *EMBO J* 5:2503–2512, 1086
- Massague J and Czech MP, The subunit structures of two distinct receptors for insulin-like growth factors I and II and their relationship to the insulin receptor. J Biol Chem 257: 5038–5045, 1982.
- LeRoith D, Clemmons D, Nissley P and Rechler MM, Insulin-like growth factors in health and disease. Ann Intern Med 116:854–862, 1992.
- 11. Sara VR and Hall K, Insulin-like growth factors and their binding proteins. *Physiol Rev* **70:**591–614, 1990.
- Clemmons DR, Role of insulin-like growth factor binding proteins in controlling IGF actions. Mol Cell Endocrinol 140:19–24, 1998.
- 13. Baserga R, Hongo A, Rubini M, Prisco M and Valentinis B, The IGF-I receptor in cell growth, transformation and apoptosis. *Biochim Biophys Acta* **1332:F**105–126, 1997.
- Rubin R and Baserga R, Insulin-like growth factor-I receptor. Its role in cell proliferation, apoptosis, and tumorigenicity. Lab Invest 73:311–331, 1995.

^{*} Brodt P, Ling H, Fallavollita L, Khatib AM and Zhang D, manuscript in preparation.

1106 P. Brodt et al.

- 15. Coppola D, Ferber A, Miura M, Sell C, D'Ambrosio C, Rubin R and Baserga R, A functional insulin-like growth factor I receptor is required for the mitogenic and transforming activities of the epidermal growth factor receptor. Mol Cell Biol 14:4588–4595, 1994.
- DeAngelis T, Ferber A and Baserga R, Insulin-like growth factor I receptor is required for the mitogenic and transforming activities of the platelet-derived growth factor receptor. J Cell Physiol 164:214–221, 1995.
- Harrington EA, Bennett MR, Fanidi A and Evan GI, c-Mycinduced apoptosis in fibroblasts is inhibited by specific cytokines. EMBO J 13:3286–3295, 1994.
- Rodriguez-Tarduchy G, Collins MK, Garcia I and Lopez-Rivas A, Insulin-like growth factor-I inhibits apoptosis in IL-3-dependent hemopoietic cells. J Immunol 149:535–540, 1992.
- 19. Resnicoff M, Abraham D, Yutanawiboonchai W, Rotman HL, Kajstura J, Rubin R, Zoltick P and Baserga R, The insulin-like growth factor I receptor protects tumor cells from apoptosis *in vivo*. Cancer Res **55:**2463–2469, 1995.
- Butler AA, Yakar S, Gewolb IH, Karas M, Okubo Y and LeRoith D, Insulin-like growth factor-I receptor signal transduction: At the interface between physiology and cell biology. Comp Biochem Physiol B Biochem Mol Biol 121:19–26, 1998.
- 21. Petley T, Graff K, Jiang W, Yang H and Florini J, Variation among cell types in the signaling pathways by which IGF-I stimulates specific cellular responses. *Horm Metab Res* 31:70–76, 1999.
- 22. White MF, The insulin signaling system and the IRS proteins. *Diabetologia* **2:**S2–17, 1997.
- Craparo A, O'Neill TJ and Gustafson TA, Non-SH2 domains within insulin receptor substrate-1 and SHC mediate their phosphotyrosine-dependent interaction with the NPEY motif of the insulin-like growth factor I receptor. J Biol Chem 270:15639–15643, 1995.
- 24. Giorgetti S, Ballotti R, Kowalski-Chauvel A, Tartare S and Van Obberghen E, The insulin and insulin-like growth factor-I receptor substrate IRS-1 associates with and activates phosphatidylinositol 3-kinase in vitro. J Biol Chem 268:7358– 7364, 1993.
- Sugimoto S, Wandless TJ, Shoelson SE, Neel BG and Walsh CT, Activation of the SH2-containing protein tyrosine phosphatase, SH-PTP2, by phosphotyrosine-containing peptides derived from insulin receptor substrate-1. J Biol Chem 269: 13614–13622, 1994.
- Sun XJ, Crimmins DL, Myers MG Jr, Miralpeix M and White MF, Pleiotropic insulin signals are engaged by multisite phosphorylation of IRS-1. Mol Cell Biol 13:7418–7428, 1993.
- Skolnik EY, Batzer A, Li N, Lee CH, Lowenstein E, Mohammadi M, Margolis B and Schlessinger J, The function of GRB2 in linking the insulin receptor to Ras signaling pathways. Science 260:1953–1955, 1993.
- 28. Valentinis B, Romano G, Peruzzi F, Morrione A, Prisco M, Soddu S, Cristofanelli B, Sacchi A and Baserga R, Growth and differentiation signals by the insulin-like growth factor 1 receptor in hemopoietic cells are mediated through different pathways. *J Biol Chem* **274:**12423–12430, 1999.
- Gual P, Baron V, Lequoy V and Van Obberghen E, Interaction of Janus kinases JAK-1 and JAK-2 with the insulin receptor and the insulin-like growth factor-1 receptor. *Endocrinology* 139:884–893, 1998.
- Zong CS, Chan J, Levy DE, Horvath C, Sadowski HB and Wang LH, Mechanism of STAT3 activation by insulin-like growth factor I receptor. J Biol Chem 275:15099–15105, 2000.
- 31. Zong CS, Zeng L, Jiang Y, Sadowski HB and Wang LH, STAT3 plays an important role in oncogenic Ros- and

- insulin-like growth factor I receptor-induced anchorage-independent growth. J Biol Chem 273:28065–28072, 1998.
- 32. Beitner-Johnson D and LeRoith D, Insulin-like growth factor-I stimulates tyrosine phosphorylation of endogenous c-Crk. J Biol Chem 270:5187–5190, 1995.
- Koval AP, Karas M, Zick Y and LeRoith D, Interplay of the proto-oncogene proteins CrkL and CrkII in insulin-like growth factor-I receptor-mediated signal transduction. J Biol Chem 273:14780–14787, 1998.
- 34. Baron V, Calleja V, Ferrari P, Alengrin F and Van Obberghen E, p125Fak focal adhesion kinase is a substrate for the insulin and insulin-like growth factor-I tyrosine kinase receptors. *J Biol Chem* **273:**7162–7168, 1998.
- 35. Casamassima A and Rozengurt E, Insulin-like growth factor I stimulates tyrosine phosphorylation of p130 (Cas), focal adhesion kinase, and paxillin. Role of phosphatidylinositol 3'-kinase and formation of a p130 (Cas). Crk complex. J Biol Chem 273:26149–26156, 1998.
- Prager D, Li HL, Yamasaki H and Melmed S, Human insulin-like growth factor I receptor internalization. Role of the juxtamembrane domain. J Biol Chem 269:11934–11937, 1994.
- Dews M, Nishimoto I and Baserga R, IGF-I receptor protection from apoptosis in cells lacking the IRS proteins. Recept Signal Transduct 7:231–240, 1997.
- 38. Li S, Ferber A, Miura M and Baserga R, Mitogenicity and transforming activity of the insulin-like growth factor-I receptor with mutations in the tyrosine kinase domain. *J Biol Chem* **269**:32558–32654, 1994.
- Hernandez-Sanchez C, Blakesley V, Kalebic T, Helman L and LeRoith D, The role of the tyrosine kinase domain of the insulin-like growth factor-I receptor in intracellular signaling, cellular proliferation and tumorigenesis. J Biol Chem 270: 29176–29181, 1995.
- 40. O'Connor R, Survival factors and apoptosis. Adv Biochem Eng Biotechnol **62:**137–166, 1998.
- 41. Miura M, Surmacz E, Burgaud JL and Baserga R, Different effects on mitogenesis and transformation of a mutation at tyrosine 1251 of the insulin-like growth factor I receptor. *J Biol Chem* **270**:22639–22644, 1995.
- 42. Blakesley VA, Koval AP, Stannard BS, Scrimgeour A and LeRoith D, Replacement of tyrosine 1251 in the carboxyl terminus of the insulin-like growth factor-I receptor disrupts the actin cytoskeleton and inhibits proliferation and anchorage-independent growth. *J Biol Chem* 273:18411–18422, 1998.
- 43. Li S, Resnicoff M and Baserga R, Effect of mutations at serines 1280–1283 on the mitogenic and transforming activities of the insulin-like growth factor I receptor. *J Biol Chem* **271**: 12254–12260, 1996.
- 44. Macaulay VM, Insulin-like growth factors and cancer. Br J Cancer 65:311–320, 1992.
- 45. Hankinson SE, Willette WC, Colditz GA, Hunter DJ, Michaud DS, Deroo B, Rosner B, Speizer FE and Pollak M, Circulating concentrations of insulin-like growth factor-I and risk of breast cancer. *Lancet* **351:**1373–1375, 1998.
- 46. Mantzoros CS, Tzonou A, Signorello LB, Stampfer M, Trichopoulos D and Adami HO, Insulin-like growth factor 1 in relation to prostate cancer and benign prostatic hyperplasia. Br J Cancer 76:1115–1118, 1997.
- 47. Ma J, Pollak MN, Giovannucci E, Chan JM, Tao Y, Hennekens CH and Stampfer MJ, Prospective study of colorectal cancer risk in men and plasma levels of insulin-like growth factor (IGF)-I and IGF-binding protein-3. *J Natl Cancer Inst* 91:620–625, 1999.
- 48. Resnicoff M, Coppola D, Sell C, Rubin R, Ferrone S and Baserga R, Growth inhibition of human melanoma cells in

- nude mice by antisense strategies to the type 1 insulin-like growth factor receptor. Cancer Res 54:4848-4850, 1994.
- Burfeind P, Chernicky CL, Rininsland F, Ilan J and Ilan J, Antisense RNA to the type I insulin-like growth factor receptor suppresses tumor growth and prevents invasion by rat prostate cancer cells in vivo. Proc Natl Acad Sci USA 93:7263–7268, 1996.
- Nakamura K, Hongo A, Kodama J, Miyagi Y, Yoshinouchi M and Kudo T, Down-regulation of the insulin-like growth factor I receptor by antisense RNA can reverse the transformed phenotype of human cervical cancer cell lines. Cancer Res 60:760–765, 2000.
- Pietrzkoski Z, Mulholland G, Gomella L, Jameson BA, Wernicke D and Baserga R. Inhibition of growth of prostatic cancer cell lines by peptide analogues of insulin-like growth factor 1. Cancer Res 53:1102–1106, 1993.
- 52. Rininsland F, Johnson TR, Chernicky CL, Schulze E, Burfeind P and Ilan J, Suppression of insulin-like growth factor type I receptor by a triple-helix strategy inhibits IGF-I transcription and tumorigenic potential of rat C6 glioblastoma cells. Proc Natl Acad Sci USA 94:5854–5859, 1997.
- 53. Baserga R, Controlling IGF-receptor function: A possible strategy for tumor therapy. *Trends Biotech* **14:**150–152, 1996.
- 54. Baserga R, The IGF-I receptor in cancer research. *Exp Cell Res* **253:**1–6, 1999.
- Long L, Nip J and Brodt P, Paracrine growth stimulation by hepatocyte-derived insulin-like growth factor-1: A regulatory mechanism for carcinoma cells metastatic to the liver. Cancer Res 54:3732–3737, 1994.
- 56. Long L, Rubin R and Brodt P, Enhanced invasion and liver colonization by lung carcinoma cells overexpressing the type 1 insulin-like growth factor receptor. Exp Cell Res 238:116– 121, 1998.
- 57. Long L, Rubin R, Baserga R and Brodt P, Loss of the metastatic phenotype in murine carcinoma cells expressing an antisense RNA to the insulin-like growth factor receptor. *Cancer Res* 55:1006–1009, 1995.
- Long L, Navab R and Brodt P, Regulation of the M_r 72,000 type IV collagenase by the type I insulin-like growth factor receptor. Cancer Res 58:3243–3247, 1998.

- 59. Burgaud JL, Resnicoff M and Baserga R, Mutant IGF-I receptors as dominant negatives for growth and transformation. *Biochem Biophys Res Commun* **214**:475–481, 1995.
- Reiss K, D'Ambrosio C, Tu X, Tu C and Baserga R, Inhibition of tumor growth by a dominant negative mutant of the insulin-like growth factor I receptor with a bystander effect. Clin Cancer Res 4:2647–2455, 1998.
- Jiang Y, Rom WN, Yie TA, Chi CX and Tchou-Wong KM, Induction of tumor suppression and glandular differentiation of A549 lung carcinoma cells by dominant-negative IGF-1 receptor. Oncogene 44:6071–6077, 1999.
- Bergeron JJ, Cruz J, Khan MN and Posner BI, Uptake of insulin and other ligands into receptor-rich endocytic components of target cells: The endosomal apparatus. Annu Rev Physiol 47:383–403, 1985.
- 63. Chow JC, Condorelli G and Smith RJ, Insulin-like growth factor-I receptor internalization regulates signaling via the Shc/mitogen-activated protein kinase pathway, but not the insulin receptor substrate-1 pathway. J Biol Chem 273:4672– 4680, 1998.
- 64. Zapf A, Hsu D and Olefsky JM, Comparison of the intracellular itineraries of insulin-like growth factor-I and insulin and their receptors in Rat-1 fibroblasts. *Endocrinology* 134:2445– 2451, 1994.
- Furlanetto RW, Receptor-mediated endocytosis and lysosomal processing of insulin-like growth factor I by mitogenically responsive cells. *Endocrinology* 122:2044–2053, 1988.
- 66. Authier R, Metioui M, Bell AW and Mort JS, Negative regulation of epidermal growth factor signaling by selective proteolytic mechanisms in the endosome mediated by cathepsin B. J Biol Chem 274:33723–33731, 1999.
- Xing R, Wu F and Mason RW, Control of breast tumor cell growth using a targeted cysteine protease inhibitor. Cancer Res 58:904–909, 1998.
- 68. Navab R, DiGulielmo GM, Bergeron JJ and Brodt P, Cysteine proteinases can regulate the malignant phenotype through their involvement in IGF-1 receptor turnover. *Proc Am Assoc Cancer Res* 40:4043, 1999.